

QUANT-BCL1/IgH MTC®

Cat.1.007

Real-Time Quantitative PCR of t(11;14) (BCL-1/IgH) MTC for diagnosis and monitoring of Mantle cell lymphoma

Introduction

Mantle cell lymphoma (MCL) is an aggressive form of non-Hodgkin (NHL) lymphoma, typically occurring in older males and usually associated with the BCL1/IgH gene rearrangement. It accounts for 4-6% of all adult NHL (1) and is usually characterized by lymphadenopathy, bone marrow (BM) and gastrointestinal tract involvement at diagnosis (2). MCL is characterized by t(11;14) translocation resulting in the juxtaposition of the BCL-1 (Cyclin D1) gene, located on chromosome 11q13, to the joining region (J) of immunoglobulin heavy chain genes (IgH), on chromosome 14q32 (BCL-1/IgH rearrangement) (Figure 1) (3,4). Nearly half of these translocations cluster within a 500 base pair region, named major translocation cluster (MTC) region, located upstream to the 5' untranslated region (5'UTR) of the BCL-1 gene on chromosome 11q13 (Figure 1). The remainder of breakpoints on chromosome 11 are widely scattered over a region of approximately 120 kb, including a minor 500 bp translocation cluster region (mtc-1) located downstream the MTC. The breakpoints on chromosome 14 occur within the 5' region of one of the six J regions of the IgH gene (5). As a consequence of the t(11;14), the BCL-1 gene, not disrupted by the translocation, undergoes the control of the em IgH enhancer (Figure 1), resulting in the constitutive overexpression of the Cyclin D1 (BCL-1) protein in lymphoma cells (6). Only translocations involving the MTC region can be revealed by a routine PCR analysis. Thus 30% to 40% of MCL cases can be shown to carry the t(11;14) translocation by PCR methods. Very recently, the quantitative evaluation of cells carrying the t(11;14) translocation, following conventional and high dose therapy, was shown to represent a powerful predictor of long-term remission in MCL patients. Therefore the possibility of measuring molecular minimal residual disease (MRD) after appropriate therapy provides a powerful tool to define subgroups of MCL patients with a significantly different prognosis (7).

Appropriate applications of the test

Molecular diagnosis of MCL is usually made either through a qualitative PCR (able to detect 30-40% of cases) or, indirectly, through detection of Cyclin D1 mRNA overexpression by REAL-time reverse transcriptase PCR. In this latter instance, Cyclin D1 overexpression needs to be referred to a B cell marker (e.g. CD19 or CD20) (8,9) and cannot be exploited for direct MRD evaluation since it does not represent a direct measurement of neoplastic clone size. Alternative methods for direct MRD evaluation such as IgH clone-specific PCR (7) are cumbersome and time consuming. In contrast, the quantitative evaluation of BCL1/IgH+ cells in tissue DNA samples (peripheral blood, bone marrow, lymph nodes, biopsies of other tissues) from patients with MTC+ MCL, as performed with the present kit, provides a rapid, reliable and reproducible tool for i) molecular diagnosis of MTC+ MCL (30-40% of MCL cases), ii) accurate and direct monitoring of minimal residual disease (MRD) of MTC+ cases, iii) early evaluation of therapeutic efficacy of a given treatment, including chemotherapy, immunotherapy, chemioimmunotherapy, high-dose therapy and radio-immunotherapy (7), iv) monitoring of tumor cell contamination in bone marrow or peripheral stem cells collected for high dose therapy with autologous hemopoietic rescue (10,11), v) early prognostic evaluation of MTC+ MCL patients (7).

Principle of assay: real-time PCR with SYBR Green® method

Applicability: On extracted and purified total DNA from peripheral blood or bone marrow lymph nodes or other tissue sample.

Number of test: 132 PCR reactions (24 samples equivalent to 72 PCR reactions + 60 PCR reactions for standard curves and controls)

Stability: 18 months if correctly stored.

Method sensitivity: the limit sensitivity of the assay (tested on the JVM-2 cell line) is of 9.0 BCL1/IgH+ cells on 90000 analyzed cells (i.e. 500 ng of tested DNA)

Kit containing and storage

Name	Storage
CD ROM 700 MB	
Software for calculation of results	
STANDARD DILUTIONS	
St1 -pMTC 9x10 ⁵ copies/2µl -Albumin 9x10 ⁴ copies/2µl	-20°C
St2 -pMTC 9x10 ⁴ copies/2µl -Albumin 9x10 ³ copies/2µl	-20°C
St3 -pMTC 9x10 ³ copies/2µl -Albumin 9x10 ² copies/2µl	-20°C
St4 -pMTC 9x10 ² copies/2µl -Albumin 9x10 ¹ copies/2µl	-20°C
St5 -pMTC 9x10 ¹ copies/2µl -Albumin 9x10 ⁰ copies/2µl	-20°C
St6 -pMTC 9x10 ⁰ copies/2µl -	-20°C
REAL-TIME PCR	
10X Albumin Primers	-20°C
10X pMTC Primers	-20°C
SYBR Green® Master Mix 2X*	+4°C
H₂O RNase/DNase-free	-20°C

Analysis of results

Cycle threshold (Ct) is defined the cycle number at which the emission intensity of reporter dye is perceptible upon background noise. In a Real-time PCR, the Ct is directly related to the copies of target-template present at the beginning of reaction. In each sample the ratio between BCL-1/IgH and Albumin is calculated using specific standard curves. For a correct normalization, the efficiencies of Albumin and BCL-1/IgH must be very similar (In a PCR reaction, the maximum efficiency correspond to a standard curve theoretical slope of -3,32)..

Below are showed the relative plots of BCL-1/IgH amplification and Standard curves (Albumin and BCL-1/IgH) dilutions.

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References

- 1) The non-Hodgkin's lymphoma classification project. *A clinical evaluation of the international lymphoma study group classification of non-Hodgkin's lymphoma*. Blood, 1997; 89: 3909-3918
- 2) Liu Y, Hernandez AM, Shibata D, and Cortopassi GA. *BCL2 translocation frequency rises with age in humans*. PNAS, 1994; 91: 8910-8914
- 3) Cleary ML, Galili N, and Sklar J. *Detection of a second t(14;18) breakpoint cluster region in human follicular lymphomas*. J.of Exp Med, 1986; 164: 315-320
- 4) Aster JC, Longtine JA. *Detection of BCL-2 rearrangements in follicular Lymphoma*. Am J Pathol, 2002; 160: 759-763.
- 5) Tsimberidou AM, Jiang J, Ford RJ, et al. *Quantitative real-time polymerase reaction for detection of circulating cells with t(14;18) in volunteer blood donors and patients with follicular lymphoma*. Leukemia & Lymphoma 2002; 43: 1589-1598
- 6) Galimberti S, Guerrini F, Morabito F, et al. *Quantitative molecular evaluation in autotransplant programs for follicular lymphoma: efficacy of in vivo purging by Rituximab*. Bone Marrow Transplant. 2003, 32:57-63.
- 7) Rambaldi A, Carlotti E, Oldani E, et al. *Quantitative PCR of bone marrow BCL2/IgH+ cells at diagnosis predicts treatment response and long-term outcome in follicular non-Hodgkin lymphoma*. Blood 2005; 105: 3428-3433.
- 8) Gascoyne RD, Adomat SA, Krajewski S, et al. *Prognostic significance of Bcl-2 protein expression and Bcl-2 gene rearrangement in diffuse aggressive non-Hodgkin's lymphoma*. Blood 1997, 90:244-51.
- 9) Mounier N, Briere J, Gisselbrecht C, et al. *Rituximab plus CHOP (R-CHOP) overcomes bcl-2--associated resistance to chemotherapy in elderly patients with diffuse large B-cell lymphoma (DLBCL)*. Blood. 2003, 101:4279-84.

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